MTQL

cobas®

Methaqualone

Order information

REF	CONTENT		Analyzer(s) on which **cobas c** pack(s) can be used
04853946 190	Abuscreen OnLine Methaqualone (200 tests)	System-ID 07 6974 6	Roche/Hitachi **cobas c** 311, **cobas c** 501/502
03304671 190	Preciset DAT Plus I calibrators CAL 1-6 (6 x 5 mL)	Codes 431-436	
03304698 190	C.f.a.s. DAT Qualitative Plus (6 x 5 mL)		
04590856 190	C.f.a.s. DAT Qualitative Plus Clinical (3 x 5 mL)	Code 699	
03312950 190	Control Set DAT I PreciPos DAT Set I (2 x 10 mL) PreciNeg DAT Set I (2 x 10 mL)		
04500873 190	Control Set DAT Clinical PreciPos DAT Clinical (2 x 10 mL) PreciNeg DAT Clinical (2 x 10 mL)		

English

System information

For **cobas c** 311/501 analyzers:

MTQLQ: ACN 199: for qualitative assay

MTQLS: ACN 200: for semiquantitative assay

MTQLC: ACN 793: for qualitative assay; using C.f.a.s. DAT Qualitative Plus Clinical

For **cobas c** 502 analyzer:

MTQLQ: ACN 8199: for qualitative assay

MTQLS: ACN 8200: for semiquantitative assay

MTQLC: ACN 8793: for qualitative assay; using C.f.a.s. DAT Qualitative Plus Clinical

Intended use

Methaqualone (MTQL) is an in vitro diagnostic test for the qualitative and semiquantitative detection of methaqualone and its metabolites in human urine on Roche/Hitachi **cobas c** systems at a cutoff concentration of 300 ng/mL. Semiquantitative test results may be obtained that permit laboratories to assess assay performance as part of a quality control program. Semiquantitative assays are intended to determine an appropriate dilution of the specimen for confirmation by a confirmatory method such as gas chromatography/mass spectrometry (GC-MS).

Methaqualone provides only a preliminary analytical test result. A more specific alternate chemical method must be used in order to obtain a confirmed analytical result. GC-MS is the preferred confirmatory method.[1] Clinical consideration and professional judgment should be applied to any drug of abuse test result, particularly when preliminary positive results are used.

Summary

Methaqualone is a quinazoline derivative that was synthesized in 1951 and found clinically effective as a sedative and hypnotic in 1956.[2] Originally introduced as a non-addictive, non-barbiturate hypnotic in 1965, both physical and psychological dependence were reported shortly after. It soon gained popularity as a street aphrodisiac, and by 1972 “luding out” (300 to 450 mg methaqualone taken with wine) was a common event among college students. This produced a feeling of indestructibility and euphoria.[3] Methaqualone is similar to all sedative hypnotics in that it often reduces inhibitions and sexual performance. The drug was removed from the U.S. market in 1984 due to its extensive misuse, however, illegal production persisted. Symptoms of methaqualone overdose range from gastrointestinal distress, drowsiness, ataxia, tingling sensations, slurred speech and muscular hyperactivity, to internal bleeding, convulsions, and coma.[4]

Methaqualone is taken orally and absorbed rapidly in the gastrointestinal tract. It is highly lipophilic and is widely distributed throughout the body. Methaqualone is bound to tissue protein at a rate of 70 to 80 % and has a slow elimination half-life which is prolonged in overdoses.[5] 72 hours after ingestion, only 40 to 50 % of a dose is excreted.[6] Metabolism occurs mainly by monohydroxylation in the hepatic microsomal enzyme system, as well as the formation of a thermally unstable N-oxide.[7] There are 5 major hydroxy metabolites, with 4’-hydroxy-methaqualone being the most predominant. Metabolites appear in urine primarily as glucuronides, with < 1 % of the parent drug excreted unchanged.[7,8] Excretion patterns of metabolites vary widely due to urine volume, individual metabolism, and tolerance to the drug.[7,8,9]

Test principle

The assay is based on the kinetic interaction of microparticles in a solution (KIMS)[10,11] as measured by changes in light transmission. In the absence of sample drug, free antibody binds to drug-microparticle conjugates causing the formation of particle aggregates. As the aggregation reaction proceeds in the absence of sample drug, the absorbance increases.

When a urine sample contains the drug in question, this drug competes with the particle-bound drug derivative for free antibody. Antibody bound to sample drug is no longer available to promote particle aggregation, and subsequent particle lattice formation is inhibited. The presence of sample drug diminishes the increasing absorbance in proportion to the concentration of drug in the sample. Sample drug content is determined relative to the value obtained for a known cutoff concentration of drug.

Reagents - working solutions

R1 Buffer; stabilizer; 0.09 % sodium azide

R2 Methaqualone antibody (goat polyclonal); buffer; bovine serum albumin; 0.09 % sodium azide

R3 Conjugated methaqualone derivative microparticles; buffer; 0.09 % sodium azide

R1 is in position B, R2 is in position C, and R3 is in position A.

Precautions and warnings

For in vitro diagnostic use.
Exercise the normal precautions required for handling all laboratory reagents.
Disposal of all waste material should be in accordance with local guidelines.
Safety data sheet available for professional user on request.

For USA: Caution: Federal law restricts this device to sale by or on the order of a physician.

Reagent handling

Ready for use

Carefully invert reagent container several times prior to use to ensure that the reagent components are mixed.

Storage and stability

Shelf life at 2-8 °C:	See expiration date on **cobas c** pack label
On-board in use and refrigerated on the analyzer:	8 weeks

Do not freeze.

Specimen collection and preparation

Only the specimens listed below were tested and found acceptable.

Urine: Collect urine samples in clean glass or plastic containers. Fresh urine specimens do not require any special handling or pretreatment, but an effort should be made to keep pipetted samples free of gross debris. Samples should be within the normal physiological pH range of 5-8. No additives or preservatives are required. It is recommended that urine specimens be stored at 2-8 °C and tested within 5 days of collection.[12]

For prolonged storage, freezing of samples is recommended.

Centrifuge highly turbid specimens before testing.

Adulteration or dilution of the sample can cause erroneous results. If adulteration is suspected, another sample should be collected. Specimen validity testing is required for specimens collected under the *Mandatory Guidelines for Federal Workplace Drug Testing Programs*[13].

CAUTION: Specimen dilutions should only be used to interpret results of Calc.? and Samp.? alarms, or when estimating concentration in preparation for GC-MS. Dilution results are not intended for patient values. Dilution procedures, when used, should be validated.

Materials provided

See "Reagents – working solutions" section for reagents.

Materials required (but not provided)

See "Order information" section

General laboratory equipment

Assay

For optimum performance of the assay follow the directions given in this document for the analyzer concerned. Refer to the appropriate operator's manual for analyzer-specific assay instructions.

The performance of applications not validated by Roche is not warranted and must be defined by the user.

Application for urine

Deselect Automatic Rerun for these applications in the Utility menu, Application screen, Range tab.

cobas c 311 **test definition**

	Semiquantitative	Qualitative
Assay type	2-Point End	2-Point End
Reaction time / Assay points	10 / 25-54	10 / 25-54
Wavelength (sub/main)	– /570 nm	– /570 nm
Reaction direction	Increase	Increase
Unit	ng/mL	mAbs

Reagent pipetting		Diluent (H_2O)
R1	46 µL	–
R2	46 µL	–
R3	40 µL	–

Sample volumes	*Sample*	*Sample dilution*	
		Sample	*Diluent (NaCl)*
Normal	5.0 µL	–	–
Decreased	5.0 µL	–	–
Increased	5.0 µL	–	–

cobas c 501/502 **test definition**

	Semiquantitative	Qualitative
Assay type	2-Point End	2-Point End
Reaction time / Assay points	10 / 37-60	10 / 37-60
Wavelength (sub/main)	– /570 nm	– /570 nm
Reaction direction	Increase	Increase
Unit	ng/mL	mAbs

Reagent pipetting		Diluent (H_2O)
R1	46 µL	–
R2	46 µL	–
R3	40 µL	–

Sample volumes	*Sample*	*Sample dilution*	
		Sample	*Diluent (NaCl)*
Normal	5.0 µL	–	–
Decreased	5.0 µL	–	–
Increased	5.0 µL	–	–

Calibration

Calibrators	*Semiquantitative application* S1-4: Preciset DAT Plus I calibrators, CAL 1-4 0, 150, 300, 600 ng/mL *Qualitative application* S1: C.f.a.s. DAT Qualitative Plus, C.f.a.s. DAT Qualitative Plus Clinical, or Preciset DAT Plus I calibrator - CAL 3 300 ng/mL The drug concentrations of the calibrators have been verified by GC-MS.
Calibration K Factor	For the qualitative application, enter the K Factor as -1000 into the Calibration menu, Status screen, Calibration Result window.
Calibration mode	*Semiquantitative application* Result Calculation Mode (RCM)[a] *Qualitative application* Linear
Calibration frequency	Full (semiquantitative) or blank (qualitative) calibration • after reagent lot change • as required following quality control procedures

a) See Results section.

Calibration interval may be extended based on acceptable verification of calibration by the laboratory.

Traceability: This method has been standardized against a primary reference method (GC-MS).

Quality control

For quality control, use control materials as listed in the "Order information" section.

In addition, other suitable control material can be used.

Drug concentrations of Control Set DAT I and Clinical have been verified by GC-MS.

The control intervals and limits should be adapted to each laboratory's individual requirements. Values obtained should fall within the defined limits. Each laboratory should establish corrective measures to be taken if values fall outside the defined limits.

Follow the applicable government regulations and local guidelines for quality control.

Results

For the qualitative assay, the cutoff calibrator is used as a reference in distinguishing between preliminary positive and negative samples. Samples producing a positive or "0" absorbance value are considered preliminary positive. Preliminary positive samples are flagged with >Test. Samples producing a negative absorbance value are considered negative. Negative samples are preceded by a minus sign.

For the semiquantitative assay, the analyzer computer constructs a calibration curve from absorbance measurements of the standards using a 4 parameter logit-log fitting function (RCM). The logit-log function fits a smooth line through the data points. The analyzer computer uses absorbance measurements of samples to calculate drug or drug metabolite concentration by interpolation of the logit-log fitting function.

NOTE: If a result of Calc.? or Samp.? alarm is obtained, review the Reaction Monitor data for the sample and compare with the Reaction Monitor data for the highest calibrator. The most likely cause is a high concentration of the analyte in the sample, in which case the absorbance value for the sample will be less than that of the highest calibrator. Make an appropriate dilution of the sample using the 0 ng/mL calibrator and rerun the sample. A normal drug-free urine may be substituted for the 0 ng/mL calibrator if the urine and procedure have been validated by the laboratory. To ensure that the sample was not over-diluted, the diluted result, prior to multiplying by the dilution factor, must be at least half the analyte cutoff value. If the diluted result falls below half the analyte cutoff value, repeat the sample with a smaller dilution. A dilution that produces a result closest to the analyte cutoff is the most accurate estimation. To estimate the preliminary positive sample's concentration, multiply the result by the appropriate dilution factor. Dilutions should only be used to interpret results of Calc.? or Samp.? alarms, or when estimating concentration in preparation for GC-MS.

Use caution when reporting results as there are various factors that influence a urine test result, such as fluid intake and other biological factors.

As with any sensitive test for drugs of abuse on automated clinical chemistry analyzers, the possibility exists for analyte carry-over from a sample with an extremely high concentration to a normal (negative) sample which immediately follows it.

Confirm all preliminary positive results by another method.

Limitations - interference

See the "Specific performance data" section of this document for information on substances tested with this assay. There is the possibility that other substances and/or factors may interfere with the test and cause erroneous results (e.g., technical or procedural errors).

A preliminary positive result with this assay indicates the presence of methaqualone and/or its metabolites in urine. It does not measure the level of intoxication.

Interfering substances were added to drug free urine at the concentration listed below. These samples were then spiked to 300 ng/mL using a methaqualone stock solution. Samples were tested in triplicate (n = 3) on a Roche/Hitachi **cobas c** 501 analyzer. The median % recoveries were calculated and are listed below.

Substance	Concentration Tested	% Methaqualone Recovery
Acetone	1 %	100
Ascorbic acid	1.5 %	104
Bilirubin	0.25 mg/mL	103
Creatinine	5 mg/mL	110
Ethanol	1 %	102
Glucose	2 %	101
Hemoglobin	7.5 g/L	86
Human albumin	0.5 %	101
Oxalic acid	2 mg/mL	112
Phenazopyridine	0.05 mg/mL	117
Sodium chloride	0.5 M	103
Sodium chloride	1 M	104
Urea	6 %	105

For diagnostic purposes, the results should always be assessed in conjunction with the patient's medical history, clinical examination and other findings.

ACTION REQUIRED

Special Wash Programming: The use of special wash steps is mandatory when certain test combinations are run together on Roche/Hitachi **cobas c** systems. The latest version of the carry-over evasion list can be found with the NaOHD-SMS-SmpCln1+2-SCCS Method Sheets. For further instructions refer to the operator's manual. **cobas c** 502 analyzer: All special wash programming necessary for avoiding carry-over is available via the **cobas** link, manual input is not required.

Where required, special wash/carry-over evasion programming must be implemented prior to reporting results with this test.

Expected values

Qualitative assay

Results of this assay distinguish preliminary positive (≥ 300 ng/mL) from negative samples only. The amount of drug detected in a preliminary positive sample cannot be estimated.

Semiquantitative assay

Results of this assay yield only approximate cumulative concentrations of the drug and its metabolites (see "Analytical specificity" section).

Specific performance data

Representative performance data on the analyzers are given below. Results obtained in individual laboratories may differ.

Precision

Precision was determined in an internal protocol by running a series of calibrator and controls (repeatability n = 20 and intermediate precision n = 100). The following results were obtained on a Roche/Hitachi **cobas c** 501 analyzer.

Semiquantitative precision

Repeatability	*Mean ng/mL*	*SD ng/mL*	*CV %*
Level 1	214	5	2.3
Level 2	295	4	1.4
Level 3	381	5	1.3

Intermediate precision	*Mean ng/mL*	*SD ng/mL*	*CV %*
Level 1	219	6	2.8
Level 2	295	5	1.5
Level 3	378	5	1.3

Qualitative precision

Cutoff (300)	Number tested	Correct results	Confidence level
0.75x	100	100	> 95 % negative reading
1.25x	100	100	> 95 % positive reading

Lower detection limit of the test

14.3 ng/mL

The lower detection limit represents the lowest measurable analyte level that can be distinguished from zero. It is calculated as the value lying 2 standard deviations above that of the lowest standard (standard 1 + 2 SD, repeatability, n = 21).

Accuracy

100 urine samples, obtained from a clinical laboratory where they screened negative in a drug test panel, were evaluated with the Methaqualone assay. 100 % of these normal urines were negative relative to the 300 ng/mL cutoff. 69 samples obtained from a clinical laboratory, where they screened preliminary positive with a commercially available immunoassay and were subsequently confirmed by GC-MS, were evaluated with the Methaqualone assay. 100 % of these samples were positive relative to the 300 ng/mL cutoff. In addition, 8 samples were diluted to a methaqualone concentration of 75-100 % of the cutoff concentration; and 7 samples were diluted to a methaqualone concentration of 100-125 % of the cutoff concentration. Data from the accuracy studies described above that fell within the near cutoff value ranges were combined with data generated from diluted positive urine samples. The following results were obtained with the Methaqualone assay on the Roche/Hitachi **cobas c** 501 analyzer relative to the GC-MS values.

Methaqualone

Methaqualone Clinical Correlation (Cutoff = 300 ng/mL)					
		Negative Samples	GC-MS values (ng/mL)		
			Near Cutoff		415-26019
			176-239	305-401	
cobas c 501 analyzer	+	0	0	10	66
	-	100	10	0	0

Analytical specificity

The specificity of this assay for structurally similar compounds was determined by generating inhibition curves for each of the compounds listed and determining the approximate quantity of each compound that is equivalent in assay reactivity to a 300 ng/mL methaqualone assay cutoff. The following results were obtained on a Roche/Hitachi **cobas c** 501 analyzer.

Compound	ng/mL Equivalent to 300 ng/mL Methaqualone	Approximate % Cross-reactivity
4'-Hydroxy-methaqualone	337	89
3'-Hydroxy-methaqualone	428	70
2'-Hydroxymethyl-methaqualone	547	55
2-Hydroxymethyl-methaqualone	720	42
6-Hydroxy-methaqualone	1825	16

Drug interference

The following compounds were added to aliquots of pooled normal human urine at a concentration of 100000 ng/mL. None of these compounds gave values in the assay that were greater than 0.16 % cross-reactivity.

Acetaminophen
Acetylsalicylic acid
Aminopyrine
Amitriptyline
Amobarbital
d-Amphetamine
l-Amphetamine
Ampicillin
Ascorbic acid
Aspartame
Atropine
Benzocaine
Benzoylecgonine
(cocaine metabolite)
Benzphetamine
Brompheniramine
Butabarbital
Caffeine
Calcium hypochlorite
Chlordiazepoxide
Chloroquine
Chlorpheniramine
Chlorpromazine
Clemastine
Cocaine
Codeine
Desipramine
Dextromethorphan
Dextropropoxyphene
Diazepam
Diphenhydramine
Diphenylhydantoin
Dopamine
Doxepin
Ecgonine
Ecgonine methyl ester
d-Ephedrine
d,l-Ephedrine
l-Ephedrine
Epinephrine
Erythromycin
Estriol
Fenoprofen
Flumazenil
Furosemide
Gentisic acid
Glutethimide
Guaiacol glycerol ether
Hydrochlorothiazide
p-Hydroxyamphetamine
Ibuprofen
Imipramine
Isoproterenol
Ketamine
Lidocaine
LSD
MDA
MDMA
Melanin
Meperidine
Methadone
d-Methamphetamine
l-Methamphetamine
Methapyrilene
Methylphenidate
Methyprylon
Morphine
Naloxone
Naltrexone
Naproxen
Niacinamide
Nordiazepam
Norethindrone
l-Norpseudoephedrine
Nortriptyline
Oxazepam
Penicillin G
Pentazocine
Pentobarbital
Phencyclidine
β-Phenethylamine
Phenobarbital
Phenothiazine
Phentermine
Phenylbutazone
d-Phenylpropanolamine
d,l-Phenylpropanolamine
Procaine
Procyclidine
Promethazine
d-Pseudoephedrine
l-Pseudoephedrine
Quinidine
Quinine
Secobarbital
Sulindac
Tetracycline
Δ^9 THC-9-carboxylic acid
Tetrahydrozoline
Thioridazine
Trifluoperazine
d,l-Trihexyphenidyl
Trimipramine
Tripelenamine
Tyramine
Verapamil
Zolpidem
Zopiclone

References

1 Karch SB, ed. Drug Abuse Handbook. Boca Raton, FL: CRC Press LLC 1998.

2 Baselt RC. Disposition of Toxic Drugs and Chemicals in Man. 7th ed. Foster City, CA: Biomedical Publications 2004.

3 Ager SA. Luding out. N Engl J Med 1972;287(1):51.

4 Abboud RT, Freemann MT, Rogers RM, et al. Methaqualone poisoning with muscular hyperactivity necessitating the use of curare. Chest 1974;65(2):204-205.

5 Nayak RK, Smyth RD, Chamberlain JH, et al. Methaqualone pharmacokinetics after single and multiple dose administration in man. J Pharmacokinet Biopharm 1974;2(2):107-121.

6 Smyth RD, Lee JK, Polk A, et al. Bioavailability of methaqualone. J Clin Pharmacol 1973;13(10):391-400.

7 Reynolds CN, Wilson K, Burnett D. Urinary excretion of methaqualone N-oxide in man. Xenobiotica 1976;6(2):113-124.

8 Wilson K, Burnett D, Oram M, et al. The kinetics of urinary excretion of the N-oxide and glucuronides of methaqualone in man. Eur J Drug Metab Pharmacokinet 1981;6(4):289-295.

9 Ericsson O, Danielsson B. Urinary excretion pattern of methaqualone metabolites in man. Drug Metab Dispos 1977;5(6):497-502.

10 Brenner C, Hui R, Passarelli J, et al. Comparison of methaqualone excretion patterns using Abuscreen ONLINE and EMIT II immunoassays and GC/MS. Forensic Sci Int 1996;79(1):31-41.

11 Armbruster DA, Schwarzhoff RH, Pierce BL, et al. Method comparison of EMIT II and ONLINE with RIA for drug screening. J Forensic Sci 1993;38:1326-1341.

12 Toxicology and Drug Testing in the Clinical Laboratory; Approved Guideline. 2nd ed. (C52-A2). Clinical and Laboratory Standards Institute 2007;27:33.

13 Mandatory Guidelines for Federal Workplace Drug Testing Programs. Fed Regist 2008 Nov 25;73:71858-71907.

A point (period/stop) is always used in this Method Sheet as the decimal separator to mark the border between the integral and the fractional parts of a decimal numeral. Separators for thousands are not used.

Symbols

Roche Diagnostics uses the following symbols and signs in addition to those listed in the ISO 15223-1 standard (for USA: see https://usdiagnostics.roche.com for definition of symbols used):

CONTENT	Contents of kit
→	Volume after reconstitution or mixing
GTIN	Global Trade Item Number

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Roche Diagnostics GmbH, Sandhofer Strasse 116, D-68305 Mannheim
www.roche.com

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Roche Diagnostics, Indianapolis, IN
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